

4. If an authorization under Standard Provisions – Reporting V.B.3 above is no longer accurate because a different individual or position has responsibility for the overall operation of the facility, a new authorization satisfying the requirements of Standard Provisions – Reporting V.B.3 above must be submitted to the Regional Water Board and State Water Board prior to or together with any reports, information, or applications, to be signed by an authorized representative (40 CFR § 122.22(c)).
5. Any person signing a document under Standard Provisions – Reporting V.B.2 or V.B.3 above shall make the following certification:

“I certify under penalty of law that this document and all attachments were prepared under my direction or supervision in accordance with a system designed to assure that qualified personnel properly gather and evaluate the information submitted. Based on my inquiry of the person or persons who manage the system or those persons directly responsible for gathering the information, the information submitted is, to the best of my knowledge and belief, true, accurate, and complete. I am aware that there are significant penalties for submitting false information, including the possibility of fine and imprisonment for knowing violations.” (40 CFR § 122.22(d)).

Any person providing the electronic signature for documents described in Standard Provisions – V.B.1, V.B.2, or V.B.3 that are submitted electronically shall meet all relevant requirements of Standard Provisions – Reporting V.B, and shall ensure that all relevant requirements of 40 CFR part 3 (Cross-Media Electronic Reporting) and 40 CFR part 127 (NPDES Electronic Reporting Requirements) are met for that submission. (40 CFR § 122.22(e)).

C. Monitoring Reports

1. Monitoring results shall be reported at the intervals specified in the MRP (Attachment E) in this Order (40 CFR § 122.41(l)(4)).
2. Monitoring results must be reported on a Discharge Monitoring Report (DMR) form or forms provided or specified by the Regional Water Board or State Water Board for reporting the results of monitoring, biosolids use, or disposal practices. As of December 21, 2016, all reports and forms must be submitted electronically to the initial recipient defined in Standard Provisions – Reporting V.J and comply with 40 CFR part 3, 40 CFR section 122.22, and 40 CFR part 127 (40 CFR § 122.41(l)(4)(i)).
3. If the Discharger monitors any pollutant more frequently than required by this Order using test procedures approved under 40 CFR part 136, or another method required for an industry-specific waste stream under 40 CFR chapter 1, subchapters N or O, the results of such monitoring shall be included in the calculation and reporting of the data submitted in the DMR or biosolids reporting form specified by the Regional Water Board (40 CFR § 122.41(l)(4)(ii)).
4. Calculations for all limitations, which require averaging of measurements, shall utilize an arithmetic mean unless otherwise specified in this Order (40 CFR § 122.41(l)(4)(iii)).

D. Compliance Schedules

Reports of compliance or noncompliance with, or any progress reports on, interim and final requirements contained in any compliance schedule of this Order, shall be submitted no later than 14 days following each schedule date (40 CFR § 122.41(l)(5)).

E. Twenty-Four Hour Reporting

1. The Discharger shall report any noncompliance which may endanger health or the environment. Any information shall be provided orally within 24 hours from the time the Discharger becomes aware of the circumstances. A report shall also be provided within five (5) days of the time the Discharger becomes aware of the circumstances. The report shall contain a description of the noncompliance and its cause; the period of noncompliance, including exact dates and times, and if the noncompliance has not been corrected, the anticipated time it is expected to continue; and steps taken or planned to reduce, eliminate, and prevent reoccurrence of the noncompliance.

For noncompliance events related to combined sewer overflows, sanitary sewer overflows, or bypass events, these reports must include the data described above (with the exception of time of discovery) as well as the type of event (i.e., combined sewer overflow, sanitary sewer overflow, or bypass event), type of overflow structure (e.g., manhole, combined sewer overflow outfall), discharge volume untreated by the treatment works treating domestic sewage, types of human health and environmental impacts of the event, and whether the noncompliance was related to wet weather.

As of December 21, 2020, all reports related to combined sewer overflows, sanitary sewer overflows, or bypass events must be submitted electronically to the initial recipient defined in Standard Provisions – Reporting V.J. The reports shall comply with 40 CFR part 3, 40 CFR section 122.22, and 40 CFR part 127. The Regional Water Board may also require the Discharger to electronically submit reports not related to combined sewer overflows, sanitary sewer overflows, or bypass events under this section. (40 CFR § 122.41(l)(6)(i)).

2. The following shall be included as information that must be reported within 24 hours:
 - a. Any unanticipated bypass that exceeds any effluent limitation in this Order (40 CFR § 122.41(l)(6)(ii)(A)).
 - b. Any upset that exceeds any effluent limitation in this Order (40 CFR § 122.41(l)(6)(ii)(B)).
3. The Regional Water Board may waive the above required written report on a case-by-case basis if an oral report has been received within 24 hours (40 CFR § 122.41(l)(6)(ii)(B)).

F. Planned Changes

The Discharger shall give notice to the Regional Water Board as soon as possible of any planned physical alterations or additions to the permitted facility. Notice is required under this provision only when (40 CFR § 122.41(l)(1)):

1. The alteration or addition to a permitted facility may meet one of the criteria for determining whether a facility is a new source in section 122.29(b) (40 CFR § 122.41(l)(1)(i)); or
2. The alteration or addition could significantly change the nature or increase the quantity of pollutants discharged. This notification applies to pollutants that are subject neither to effluent limitations in this Order nor to notification requirements under section

122.42(a)(1) (see Additional Provisions—Notification Levels VII.A.1). (40 CFR § 122.41(l)(1)(ii)).

3. The alteration or addition results in a significant change in the Discharger's sludge use or disposal practices, and such alteration, addition, or change may justify the application of permit conditions that are different or absent in the existing permit, including notification and additional use or disposal sites not reported during the permit application process or not reported pursuant to an approved land application plan (40 CFR 122.41(l)(1)(iii)).

G. Anticipated Noncompliance

The Discharger shall give advance notice to the Regional Water Board of any planned changes in the permitted facility or activity that may result in noncompliance with this Order's requirements (40 CFR § 122.41(l)(2)).

H. Other Noncompliance

The Discharger shall report all instances of noncompliance not reported under Standard Provisions – Reporting V.C, V.D, and V.E above at the time monitoring reports are submitted. The reports shall contain the information listed in Standard Provision – Reporting V.E above. For noncompliance events related to combined sewer overflows, sanitary sewer overflows, or bypass events, these reports shall contain the information described in Standard Provision – Reporting V.E and the applicable required data in appendix A to 40 CFR part 127. The Regional Water Board may also require the Discharger to electronically submit reports not related to combined sewer overflows, sanitary sewer overflows, or bypass events under this section (40 CFR § 122.41(l)(7)).

I. Other Information

When the Discharger becomes aware that it failed to submit any relevant facts in a permit application, or submitted incorrect information in a permit application or in any report to the Regional Water Board, State Water Board, or USEPA, the Discharger shall promptly submit such facts or information (40 CFR § 122.41(l)(8)).

J. Initial Recipient for Electronic Reporting Data

The owner, operator, or the duly authorized representative is required to electronically submit NPDES information specified in appendix A to 40 CFR part 127 to the initial recipient defined in 40 CFR section 127.2(b). USEPA will identify and publish the list of initial recipients on its website and in the Federal Register, by state and by NPDES data group [see 40 CFR section 127.2(c)]. USEPA will update and maintain this listing (40 CFR § 122.41(l)(9)).

VI. STANDARD PROVISIONS – ENFORCEMENT

- A. The Regional Water Board is authorized to enforce the terms of this permit under several provisions of the CWC, including, but not limited to, sections 13268, 13385, 13386, and 13387.
- B. The CWA provides that any person who violates section 301, 302, 306, 307, 308, 318 or 405 of the CWA, or any permit condition or limitation implementing any such sections in a permit issued under section 402, or any requirement imposed in a pretreatment program approved under sections 402(a)(3) or 402(b)(8) of the CWA, is subject to a civil penalty not to exceed \$25,000 per day for each violation. The CWA provides that any person who negligently violates sections 301, 302, 306, 307, 308, 318, or 405 of the CWA, or any condition or limitation implementing any of such sections in a permit issued under section 402 of the CWA, or any requirement imposed in a pretreatment program approved under section 402(a)(3) or 402(b)(8) of the CWA, is subject to criminal penalties of \$2,500 to \$25,000 per day of violation, or imprisonment of not more than one year, or both. In the case of a second or subsequent conviction for a negligent violation, a person shall be subject to criminal penalties

of not more than \$50,000 per day of violation, or by imprisonment of not more than two years, or both. Any person who knowingly violates such conditions or limitations is subject to criminal penalties of \$5,000 to \$50,000 per day of violation, or imprisonment for not more than three years, or both. In the case of a second or subsequent conviction for a knowing violation, a person shall be subject to criminal penalties of not more than \$100,000 per day of violation, or imprisonment of not more than 6 years, or both. Any person who *knowingly* violates section 301, 302, 303, 306, 307, 308, 318 or 405 of the CWA, or any permit condition or limitation implementing any of such sections in a permit issued under section 402 of the CWA, and who knows at that time that he thereby places another person in imminent danger of death or serious bodily injury, shall, upon conviction, be subject to a fine of not more than \$250,000 or imprisonment of not more than 15 years, or both. In the case of a second or subsequent conviction for a knowing endangerment violation, a person shall be subject to a fine of not more than \$500,000 or by imprisonment of not more than 30 years, or both. An organization, as defined in section 309(c)(3)(B)(iii) of the CWA, shall, upon conviction of violating the imminent danger provision, be subject to a fine of not more than \$1,000,000 and can be fined up to \$2,000,000 for second or subsequent convictions (40 CFR § 122.41(a)(2); CWC section 13385 and 13387).

- C. Any person may be assessed an administrative penalty by the Administrator of USEPA, or an administrative civil liability by the Regional Water Board, or State Water Board for violating section 301, 302, 306, 307, 308, 318 or 405 of this CWA, or any permit condition or limitation implementing any of such sections in a permit issued under section 402 of the CWA. Administrative penalties for Class I violations are not to exceed \$10,000 per violation, with the maximum amount of any Class I penalty assessed not to exceed \$25,000. Penalties for Class II violations are not to exceed \$10,000 per day for each day during which the violation continues, with the maximum amount of any Class II penalty not to exceed \$125,000 (40 CFR § 122.41(a)(3)).
- D. The CWA provides that any person who falsifies, tampers with, or knowingly renders inaccurate any monitoring device or method required to be maintained under this permit shall, upon conviction, be punished by a fine of not more than \$10,000, or by imprisonment for not more than two years, or both. If a conviction of a person is for a violation committed after a first conviction of such person under this paragraph, punishment is a fine of not more than \$20,000 per day of violation, or by imprisonment of not more than four years, or both (40 CFR § 122.41(j)(5)).
- E. The CWA provides that any person who knowingly makes any false statement, representation, or certification in any record or other document submitted or required to be maintained under this permit, including monitoring reports or reports of compliance or non-compliance shall, upon conviction, be punished by a fine of not more than \$10,000 per violation, or by imprisonment for not more than six months per violation, or by both (40 CFR § 122.41(k)(2)).

VII. ADDITIONAL PROVISIONS – NOTIFICATION LEVELS

A. Publicly-Owned Treatment Works (POTWs)

All POTWs shall provide adequate notice to the Regional Water Board of the following (40 CFR § 122.42(b)):

1. Any new introduction of pollutants into the POTW from an indirect discharger that would be subject to sections 301 or 306 of the CWA if it were directly discharging those pollutants (40 CFR § 122.42(b)(1)); and
2. Any substantial change in the volume or character of pollutants being introduced into that POTW by a source introducing pollutants into the POTW at the time of adoption of the Order (40 CFR § 122.42(b)(2)).

3. Adequate notice shall include information on the quality and quantity of effluent introduced into the POTW as well as any anticipated impact of the change on the quantity or quality of effluent to be discharged from the POTW (40 CFR § 122.42(b)(3)).

E.

ATTACHMENT E – MONITORING AND REPORTING PROGRAM

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ATTACHMENT E – MONITORING AND REPORTING PROGRAM (MRP CI 2022)

Section 308 of the federal Clean Water Act (CWA) and sections 122.41(h), (j)-(l), 122.44(i), and 122.48 of title 40 of the Code of Federal Regulations (40 CFR) require that all NPDES permits specify monitoring and reporting requirements. California Water Code (CWC) sections 13267 and 13383 also authorize the Regional Water Board to establish monitoring, inspection, entry, reporting, and recordkeeping requirements. This MRP establishes monitoring, reporting, and recordkeeping requirements that implement the federal and California laws and/or regulations.

I. GENERAL MONITORING PROVISIONS

- A.** All samples shall be representative of the waste discharge under conditions of peak load. Quarterly effluent analyses shall be performed during the first quarter (January, February, and March), the second quarter (April, May, and June), the third quarter (July, August, and September), and the fourth quarter (October, November, and December). Semiannual analyses shall be performed during the first quarter (January, February, and March) and third quarter (July, August, and September). Annual analyses shall be performed during the third quarter (July, August, and September). Should there be instances when monitoring could not be performed during these specified months, the Discharger must notify the Regional Water Board and state the reason why monitoring could not be conducted, and obtain approval from the Executive Officer for an alternate schedule. Results of quarterly, semiannual, and annual analyses shall be reported by the due date specified in Table E-11 of the MRP.
- B.** Pollutants shall be analyzed using the analytical methods described in 40 CFR § 136.3, 136.4, and 136.5; or where no methods are specified for a given pollutant, by methods approved by this Regional Water Board or the State Water Board. Laboratories analyzing effluent samples and receiving water samples shall be certified by the State Water Resources Control Board, Division of Drinking Water (DDW) Environmental Laboratory Accreditation Program (ELAP) or approved by the Executive Officer and must include quality assurance/quality control (QA/QC) data in their reports. A copy of the laboratory certification shall be provided in the Annual Report due to the Regional Water Board each time a new certification and/or renewal of the certification is obtained from ELAP.
- C.** Water/wastewater samples must be analyzed within allowable holding time limits as specified in 40 CFR § 136.3. All QA/QC analyses must be run on the same dates that samples are actually analyzed. The Discharger shall retain the QA/QC documentation in its files and make available for inspection and/or submit this documentation when requested by the Regional Water Board. Proper chain of custody procedures must be followed and a copy of this documentation shall be submitted with the monthly report.
- D.** The Discharger shall calibrate and perform maintenance procedures on all monitoring instruments to insure accuracy of measurements, or shall ensure that both equipment activities will be conducted.
- E.** For any analyses performed for which no procedure is specified in the United States Environmental Protection Agency (USEPA) guidelines, or in the MRP, the constituent or parameter analyzed and method or procedure used must be specified in the monitoring report.
- F.** Each monitoring report must affirm in writing that “all analyses were conducted at a laboratory certified for such analyses under the Environmental Laboratory Accreditation Program (ELAP), or approved by the Executive Officer and in accordance with current USEPA guideline procedures or as specified in this monitoring and reporting program.”

- G. The monitoring report shall specify the USEPA analytical method used, the Method Detection Limit (MDL), and the Reporting Level (RL) [the applicable Minimum Level (ML) or Reported Minimum Level (RML)] for each pollutant. The MLs are those published by the State Water Board in Appendix II of the 2015 Ocean Plan. The ML represents the lowest quantifiable concentration in a sample based on the proper application of all method-based analytical procedures and the absence of any matrix interference. When all specific analytical steps are followed and after appropriate application of method specific factors, the ML also represents the lowest standard in the calibration curve for that specific analytical technique. When there is deviation from the analytical method for dilution or concentration of samples, other factors are applied to the ML depending on the sample preparation. The resulting value is the reported Minimum Level.
- H. The Discharger shall select the analytical method that provides an ML lower than the effluent limitation or performance goal established for a given parameter or where no such requirement exists, the lowest applicable water quality objective in the Ocean Plan. If the effluent limitation, performance goal, or the lowest applicable water quality objective is lower than all the MLs in Appendix II of the 2015 Ocean Plan, the Discharger must select the method with the lowest ML for compliance purposes. The Discharger shall include in the annual summary reports a list of the analytical methods and MLs employed for each test.
- I. The Discharger shall instruct its laboratories to establish calibration standards so that the ML (or its equivalent if there is differential treatment of samples relative to calibration standards) is the lower calibration standard. At no time is the Discharger to use analytical data derived from extrapolation beyond the lowest point of the calibration curve.
- J. The Discharger shall develop and maintain a record of all spills or bypasses of raw or partially treated sewage from its collection system or treatment plant according to the requirements in the WDR section of this Order. This record shall be made available to the Regional Water Board upon request and a spill summary shall be included in the annual summary report.
- K. If the Discharger samples and performs analyses (other than for process/operational control, startup, research, or equipment testing) any influent, effluent, or receiving water constituent more frequently than required by this Order using approved analytical methods, the results of those analyses shall be included in the monitoring report. These results shall be reflected in the calculation of the average (or median) used in demonstrating compliance with limitations set forth in this Order.
- L. For all bacterial analyses, sample dilutions should be performed so the expected range of values is bracketed (for example, with multiple tube fermentation method or membrane filtration method, 2 to 16,000 per 100 mL for total and fecal coliforms, at a minimum; and 1 to 1000 per 100 mL for *Enterococcus*). The detection methods used for each analysis shall be reported with the results of the analyses.
 - 1. Detection methods used for coliforms (total and fecal) shall be those presented in Table 1A of 40 CFR § 136, unless alternate methods have been approved in advance by the USEPA pursuant to 40 CFR § 136.
 - 2. Detection methods for *Escherichia coli* shall be those presented in Table 1A of 40 CFR § 136 or in the USEPA publication EPA 600/4-85/076, Test Methods for *Escherichia coli* and enterococci in Water By Membrane Filter Procedure, or any improved method determined by the Regional Water Board to be appropriate.
- M. All receiving and ambient water monitoring conducted in compliance with the MRP must be comparable with the Quality Assurance requirements of the Surface Water Ambient Monitoring Program (SWAMP). The Discharger shall submit all receiving water monitoring data in accordance with the California Environmental Exchange Network (CEDEN).

- N. NPDES compliance monitoring focuses on the effects of a specific point source discharge. Generally, it is not designed to assess impacts from other sources of pollution (e.g., nonpoint source runoff, aerial fallout) or to evaluate the current status of important ecological resources in the water body. The scale of existing compliance monitoring programs does not match the spatial and, to some extent, temporal boundaries of the important physical and biological processes in the ocean. In addition, the spatial coverage provided by compliance monitoring programs is less than ten percent of the nearshore ocean environment. Better technical information is needed about status and trends in ocean waters to guide management and regulatory decisions, to verify the effectiveness of existing programs, and to shape policy on marine environmental protection.
- O. The Regional Water Board and USEPA, working with other groups, have developed a comprehensive basis for effluent and receiving water monitoring appropriate to large publicly owned treatment works (POTWs) discharging to waters of the Southern California Bight. This effort has culminated in the publication by the Southern California Coastal Water Research Project (SCCWRP) of the Model Monitoring Program guidance document (Schiff, K.C., J.S. Brown and S.B. Weisberg. 2001. *Model Monitoring Program for Large Ocean Dischargers in Southern California*. SCCWRP Tech. Rep. #357. Southern California Coastal Water Research Project, Westminster, CA. 101 pp.). This guidance provides the principles, framework and recommended design for effluent and receiving water monitoring elements that have guided development of the monitoring program described below.
- P. In July 2000, the Santa Monica Bay Restoration Project (SMBRP) published "An Assessment of the Compliance Monitoring System in Santa Monica Bay" to set forth recommendations and priorities for compliance monitoring in Santa Monica Bay. This report reasoned that a reduced level of receiving water monitoring is justified for large POTWs discharging to Santa Monica Bay due to improvements in effluent quality and associated decreases in receiving water impacts. Like the Model Monitoring Plan developed by SCCWRP, SMBRP recommendations are focused on providing answers to management questions and allowing a reduction in POTW receiving water monitoring where discharge effects are well understood. Although the OWTS does not discharge into Santa Monica Bay, receiving water monitoring since 1999 has documented that marine conditions are consistent or superior at the City's outfall. OWTP has also participated in the Central Bight Water Quality Consortium (CALCOFI study), implemented by the NOAA and NMFS, to identify marine trends which extend beyond Santa Monica Bay. The monitoring plan set forth here has been guided by SMBRP recommendations.
- Q. The conceptual framework for the Model Monitoring Program has three components that comprise a range of spatial and temporal scales: (1) core monitoring; (2) regional monitoring; and (3) special studies.
1. Core monitoring is local in nature and focused on monitoring trends in quality and effects of the point source discharge. This includes effluent monitoring as well as some aspects of receiving water monitoring. In the monitoring program described below, these core components are typically referred to as local monitoring.
 2. Regional monitoring is focused on questions that are best answered by a region-wide approach that incorporates coordinated survey design and sampling techniques. The major objective of regional monitoring is to collect information required to assess how safe it is to swim in the ocean, how safe it is to eat seafood from the ocean, and whether the marine ecosystem is being protected. Key components of regional monitoring include elements to address pollutant mass emission estimations, public health concerns, monitoring of trends in natural resources, assessment of regional impacts from all contaminant sources, and protection of beneficial uses. The final design of regional monitoring programs is developed by means of steering committees and technical

committees comprised of participating agencies and organizations and is not specified in this Order. Instead, for each regional component, the degree and nature of participation of the Discharger is specified. For this Order, these levels of effort are based upon past participation of the Discharger in regional monitoring programs.

3. Special studies are focused on refined questions regarding specific effects or development of monitoring techniques and are anticipated to be of short duration and/or small scale, although multiyear studies also may be needed. Questions regarding effluent or receiving water quality, discharge impacts, ocean processes in the area of the discharge, or development of techniques for monitoring the same, arising out of the results of core or regional monitoring, may be pursued through special studies. These studies are by nature ad hoc and cannot be typically anticipated in advance of the five-year permit cycle.

The Discharger and the Regional Water Board shall consult annually to determine the need for special studies. Each year, the Discharger shall submit proposals for any proposed special studies to the Regional Water Board by December 31st for the following year's monitoring effort (July through June). The following year, detailed scopes of work for proposals, including reporting schedules, shall be presented by the Discharger at a Spring Regional Water Board meeting, to obtain the Regional Water Board approval and to inform the public. Upon approval by the Regional Water Board, the Discharger shall implement its special study or studies.

R. Discharger participation in regional monitoring programs is required as a condition of this Order.

1. Central Bight Water Quality Cooperative Program – The City of Oxnard has participated in coordinated monitoring conducted by the Orange County Sanitation District, County Sanitation Districts of Los Angeles County, and the City of Los Angeles through appropriate agencies for water quality monitoring as part of the federally funded Southern California Coastal Ocean Observing System (SCCOOS), which contributes to the national U.S. Integrated Ocean Observing System. This regionally coordinated survey provided integrated water quality surveys on a quarterly basis. These surveys cover 200 kilometers of coast in Ventura, Los Angeles, and Orange Counties, from the nearshore to approximately 10 kilometers offshore. This cooperative program contributes to a regional understanding of seasonal patterns in nearshore water column structure and provides context for determining the significance and potential causes of locally observed patterns in the area of wastewater outfalls. The study is also coordinated with the California Cooperative Oceanic Fisheries Investigations (CalCOFI) implemented by the National Oceanographic Atmospheric Administration (NOAA) and the California Department of Fish and Wildlife. Although monitoring as part of this program is no longer required, the City of Oxnard shall participate in planning or analysis activities as necessary to interpret the data collected over the previous 20 years.
2. The Central Region Kelp Survey Consortium was established with the support of the Regional Water Board to conduct regional kelp bed monitoring. This program is designed to require ocean dischargers in the Regional Water Board's jurisdiction to undertake a collaborative program, which include participation by Orange County ocean dischargers, to monitor kelp beds in the Southern California Bight, patterned after the successful program implemented by the San Diego Regional Water Board since 1985. Data collected in this regional survey is used to assess status and trends in kelp bed health and spatial extent. The regional nature of the survey allows the status of beds local to specific dischargers to be compared to regional trends. The regional kelp monitoring survey was initiated during 2003.

3. Southern California Bight Studies – Every five years SCCWRP coordinates regional monitoring within the Southern California Bight and compiles monitoring data collected by the dischargers and other participating entities. The fifth regional monitoring program (Bight '13) occurred primarily during summer 2013. The sixth regional monitoring program (Bight '18) is taking place during 2018. The next (seventh) regional monitoring program (Bight '23) is expected to take place during the summer of 2023. While participation in regional monitoring programs is required under this Order, revisions to the Discharger's monitoring program at the direction of the Regional Water Board may be necessary to accomplish the goals of regional monitoring or to allow the performance of special studies to investigate regional or site-specific water issues of concern. These revisions may include a reduction or increase in the number of parameters to be monitored, the frequency of monitoring, or the number and size of samples to be collected. Such changes were and may be authorized by the Regional Water Board Executive Officer after written notification to the Discharger.
- S. This monitoring program for OWTP is comprised of requirements to demonstrate compliance with the conditions of the NPDES permit, ensure compliance with State water quality standards, and mandate participation in regional monitoring and/or area-wide studies.
- T. Laboratory Certification. Laboratories analyzing monitoring samples shall be certified by the State Water Resources Control Board (State Water Board), in accordance with the provision of Water Code section 13176, and must include quality assurance/quality control data with their reports.
- U. The Discharger shall ensure that the results of the Discharge Monitoring Report-Quality Assurance (DMR-QA) Study or the most recent Water Pollution Performance Evaluation Study are submitted annually to the State Water Board at the following address:

State Water Resources Control Board
Quality Assurance Program Officer
Office of Information Management and Analysis
1001 I Street, Sacramento, CA 95814

II. MONITORING LOCATIONS

- A. The Discharger shall establish the following monitoring locations to demonstrate compliance with the effluent limitations, discharge specifications, and other requirements in this Order:

Table E-1. Influent and Effluent Monitoring

Discharge Point Name	Monitoring Location Name	Monitoring Location Description
Influent Monitoring Station		
	INF-001	Sampling station shall be established at point of inflow to the sewage treatment plant where representative samples of the influent can be obtained.
Effluent Monitoring Stations		
001	EFF-001A	The effluent sampling station shall be located as seen in Attachment C-1, where a representative sample of the secondary-treated effluent before mixing with the brine waste can be obtained.
	EFF-001B	The effluent sampling station shall be located downstream of any in-plant return flows and after the brine waste produced from the AWWPF has commingled with the final secondary effluent, where representative samples of the effluent can be obtained.

Table E-2. Local Receiving Water Quality Monitoring Station Locations

Core Receiving Water Monitoring Stations (Attachment B-4)					
Monitoring Point Name	Monitoring Location Description				
Water Column	A001	B001	C001	D001	E001
	A002	B002	C002	D002	E002
	A003	B003	C003	D003	E003
	A004	B004	C004	D004	E004
	A005	B005	C005	D005	E005
Latitude	34.138081	34.136325	34.132179	34.126674	34.120332
	34.132012	34.129138	34.124922	34.119997	34.114238
	34.125161	34.122454	34.117716	34.113029	34.107623
	34.117883	34.115150	34.110515	34.105505	34.100469
	34.110413	34.107873	34.103261	34.098363	34.093656
Longitude	-119.212132	-119.201523	-119.192064	-119.183273	-119.175354
	-119.218126	-119.208035	-119.198634	-119.189477	-119.181209
	-119.224458	-119.214308	-119.205188	-119.195943	-119.186815
	-119.231060	-119.220927	-119.211638	-119.202504	-119.193520
	-119.237707	-119.227089	-119.218082	-119.208956	-119.199672
Station Depth (m)	13	11	12	11	11
	32	15	16	16	16
	143	21	22	22	22
	219	64	28	29	31
	82	258	35	37	38
Distance from Outfall/Transect length (m)	2,000	1,000	0	1,000	2,000

Table E-3. Receiving Water Benthic, Trawl, Shoreline and Mussel Monitoring Station Locations

Monitoring Point Name	Monitoring Location Description						
Receiving Water Benthic Monitoring Stations (Attachment B-5)							
Benthic Monitoring Station	RWS-001	RWS-002	RWS_003	RWS-004	RWS-005	RWS-006	RWS-007
Latitude	34.13047	34.12655	34.12568	34.12578	34.12502	34.12328	34.1075
Longitude	-119.20887	-119.20025	-119.19831	-119.1979	-119.19643	-119.1934	-119.17017
Station Depth (m)	15.0	15.0	15.3	15.0	15.3	15.3	15.3
Dist. From Outfall Transect (m)	1000	150	18	18	150	500	4000
Bagged Mussel Stations (Attachment B-6)							
SS1	2.0 miles southeast of outfall			34.107496		-119.170076	
SS2	outfall			34.124061		-119.199514	
SS3	0.8 miles northwest of outfall			34.130546		-119.208852	

Monitoring Point Name	Monitoring Location Description		
Receiving Water Trawl Stations (Attachment B-7)			
Trawl Station	RWT-001	RWT-002	RWT-003
Latitude	34.13177	34.12308	34.11773
Longitude	-119.2063	-119.1906	-119.1833
Station Depth (m)	15.6	15.6	15.6
Distance from Outfall Transect (m)	380	380	4000
Monitoring Point Name	Monitoring Location Description		
Shoreline Bacteriological Monitoring Stations (Attachment B-8)			
Ventura County ID	Location	Latitude	Longitude
35000	Hollywood Beach, Los Robles St	34.1625	-119.2300
37000	Channel Islands Harbor Beach	34.1594	-119.2219
38000	Silverstrand Beach, San Nicholas Ave	34.1572	-119.2253
39000	Silverstrand Beach, Santa Paula Ave	34.1525	-119.2197
40000	Silverstrand Beach, Sawtell, Ave	34.1475	-119.2164
41000	Port Hueneme Beach Park	34.1417	-119.1944
42000	Ormond Beach, J Street Drain	34.1389	-119.1889
43000	Ormond Beach, Industrial Drain	34.1358	-119.1842
44000	Ormond Beach, Arnold Rd	34.1197	-119.1600

The North latitude and West longitude information in Table E-1 through E-3 are approximate for administrative purposes.

III. INFLUENT MONITORING REQUIREMENTS

Influent monitoring is required to determine compliance with NPDES permit conditions, to assess treatment plant performance and to assess effectiveness of the Pretreatment Program.

A. Monitoring Location INF-001

1. The Discharger shall monitor influent to the facility at INF-001 as follows. If more than one analytical test method is listed for a given parameter, the Discharger must select from the listed methods and corresponding Minimum Level:

Table E-4. Influent Monitoring

Parameter	Units	Sample Type	Minimum Sampling Frequency	Required Analytical Test Method
Flow	MGD	Recorder/totalizer	continuous	1
Biochemical Oxygen Demand (BOD ₅ 20°C)	mg/L	24-hr composite	weekly	2
Total Suspended Solids (TSS)	mg/L	24-hr composite	weekly	2
pH	pH units	grab	weekly	2
Oil and Grease	mg/L	grab ³	weekly	2
Radioactivity (including gross alpha, gross, beta, combined radium-226 & radium-228, tritium, strontium-90 and uranium)	pCi/L	24-hr composite	quarterly	4
Benzidine	µg/L	grab	semiannually	2
Polychlorinated Biphenyls (PCBs)	µg/L	24-hr composite	semiannually	2
TCDD Equivalents	µg/L	24-hr composite	semiannually	5
Remaining pollutants in Table B of the 2015 Ocean Plan (excluding residual chlorine, chronic toxicity)	µg/L	grab	semiannually	2

IV. EFFLUENT MONITORING REQUIREMENTS

Effluent monitoring is required to determine compliance with NPDES permit conditions and water quality standards; assess and improve plant performance, and identify operational problems; provide information on wastewater characteristics and flows for use in interpreting water quality and biological data; and to conduct reasonable potential analyses for toxic pollutants.

A. Monitoring Location EFF-001A and EFF-001B

The Discharger shall monitor at effluent monitoring locations EFF-001A or EFF-001B for all parameters as follows. The samples for BOD and percent removal of BOD, TSS and percent removal of TSS and bacteria shall be collected at monitoring location EFF-001A and all remaining samples shall be collected at EFF-001B. Compliance with TSS percent removal at EFF-001B and Ocean Plan requirements (75%) is demonstrated by compliance at EFF-001A (85%).

¹When continuous monitoring of flow is required, total daily flow, monthly average flow, and instantaneous peak daily flow (24-hour basis) shall be reported. Actual monitored flow shall be reported (not design capacity).

² Pollutants shall be analyzed using the analytical methods described in 40 CFR § 136; or where no methods are specified for a given pollutant, by methods approved by this Regional Water Board, the State Water Board, and USEPA Region 9. For any pollutant whose effluent limitation is lower than all the MLs specified in Appendix II of the Ocean Plan, the analytical method with the lowest ML must be selected.

³ Oil and grease monitoring shall consist of a single grab sample at peak flow over a 24-hour period.

⁴ Analyze these radiochemicals by the following USEPA methods: method 900.0 for gross alpha and gross beta, method 903.0 or 903.1 for radium-226, method 904.0 for radium-228, method 906.0 for tritium, method 905.0 for strontium-90, and method 908.0 for uranium. Analysis for combined radium-226 & 228 shall be conducted only if gross alpha and gross beta results for the same sample exceed 15 pCi/L or 50 pCi/L, respectively. If radium-226 & 228 exceeds 5 pCi/L, then analyze for tritium, strontium-90, and uranium.

⁵ USEPA Method 1613 shall be used to analyze TCDD equivalents.

If more than one analytical test method is listed for a given parameter, the Discharger must select from the listed methods and corresponding Minimum Level:

Table E-5. Effluent Monitoring

Parameter	Units	Sample Type ⁶	Minimum Sampling Frequency ⁷	Required Analytical Test Method
Flow	MGD	recorder/totalizer	continuous	1
BOD ₅ 20°C ⁸	mg/L	24-hr composite	weekly	2
Suspended solids ⁸	mg/L	24-hr composite	weekly	2
pH	pH units	grab	weekly	2
Oil and grease	mg/L	grab ⁹	weekly	2
Temperature	°F	grab	weekly	2
Settleable solids	ml/L	grab	weekly	2
Total chlorine residual	mg/L	recorder/totalizer	continuous	2
Turbidity	NTU	recorder/totalizer	continuous	2
Total coliform ⁸	CFU/100 ml or MPN/100 ml	grab	daily	2
<i>Enterococcus</i> ⁸	CFU/100 ml or MPN/100 ml	grab	5 times/month	2
Fecal coliform ⁸	CFU/100 ml or MPN/100 ml	grab	5 times/month	2
Ammonia nitrogen	mg/L	24-hr composite	monthly	2
Toxicity, chronic ¹⁰	Pass or Fail, % Effect (TST)	24-hr composite	monthly	2
Nitrate nitrogen	mg/L	24-hr composite	monthly	2
Nitrite nitrogen	mg/L	24-hr composite	monthly	2
Organic nitrogen	mg/L	24-hr composite	monthly	2
Radioactivity	pCi/L	24-hr composite	quarterly	11

⁶ For discharge durations of less than 24 hours, individual grab samples may be substituted. A grab sample is an individual sample collected in less than 15 minutes.

⁷ For the influent and effluent, weekly and monthly sampling shall be arranged so that each day, Monday through Friday, is represented.

⁸ Sampling for BOD, TSS and bacteria is at EFF-001A. Weekly and Monthly Average BOD may be calculated from daily measurements. If the result of the weekly BOD analysis yields a value greater than the average monthly effluent limitation (AMEL), the frequency of analysis shall be increased to daily within one week of knowledge of the test result for at least 30 days and until compliance with the average weekly effluent limitation (AWEL) and AMEL BOD limits is demonstrated; after which the frequency shall revert to weekly.

⁹ Oil and grease monitoring shall consist of a single grab sample at peak flow over a 24-hour period.

¹⁰ Whole effluent toxicity monitoring is required for Discharge Point 001, using samples collected at EFF-001B, see section V. Whole Effluent Toxicity Testing Requirements.

¹¹ Analyze these radiochemicals by the following USEPA methods: method 900.0 for gross alpha and gross beta, method 903.0 or 903.1 for radium-226, method 904.0 for radium-228, method 906.0 for tritium, method 905.0 for strontium-90, and method 908.0 for uranium. Analysis for combined radium-226 & 228 shall be conducted

Parameter	Units	Sample Type ⁶	Minimum Sampling Frequency ⁷	Required Analytical Test Method
(Including gross alpha, gross beta, combined radium-226 and radium-228, tritium, strontium-90 and uranium)				
Arsenic	µg/L	24-hr composite	semiannually	2
Cadmium	µg/L	24-hr composite	semiannually	2
Chromium (VI)	µg/L	Grab	semiannually	2
Copper	µg/L	24-hr composite	semiannually	2
Lead	µg/L	24-hr composite	semiannually	2
Mercury	µg/L	24-hr composite	semiannually	2
Nickel	µg/L	24-hr composite	semiannually	2
Selenium	µg/L	24-hr composite	semiannually	2
Silver	µg/L	24-hr composite	semiannually	2
Zinc	µg/L	24-hr composite	semiannually	2
Cyanide	µg/L	grab	semiannually	2
Phenolic compounds (non-chlorinated) ¹²	µg/L	24-hr composite	semiannually	2
Phenolic compounds (chlorinated) ¹²	µg/L	24-hr composite	semiannually	2
Endosulfan	µg/L	24-hr composite	semiannually	2
Endrin	µg/L	24-hr composite	semiannually	2
HCH ¹²	µg/L	24-hr composite	semiannually	2
Acrolein	µg/L	grab	semiannually	2
Antimony	µg/L	24-hr composite	semiannually	2
Bis(2-chloroethoxy) methane	µg/L	24-hr composite	semiannually	2
Bis(2-chloroisopropyl) ether	µg/L	24-hr composite	semiannually	2
Chlorobenzene	µg/L	grab	semiannually	2
Chromium (III)	µg/L	grab	semiannually	2
Di-n-butyl phthalate	µg/L	24-hr composite	semiannually	2
Dichlorobenzenes	µg/L	24-hr composite	semiannually	2
Diethyl Phthalate	µg/L	24-hr composite	semiannually	2

only if gross alpha and gross beta results for the same sample exceed 15 pCi/L or 50 pCi/L, respectively. If radium-226 & 228 exceeds the stipulated criteria, then analyze for tritium, strontium-90, and uranium.

¹² See Attachment A for definition of terms

Parameter	Units	Sample Type ⁶	Minimum Sampling Frequency ⁷	Required Analytical Test Method
Dimethyl Phthalate	µg/L	24-hr composite	semiannually	2
4,6-dinitro-2-methylphenol	µg/L	24-hr composite	semiannually	2
2,4-dinitrophenol	µg/L	24-hr composite	semiannually	2
Ethylbenzene	µg/L	grab	semiannually	2
Fluoranthene	µg/L	24-hr composite	semiannually	2
Hexachlorocyclopentadiene	µg/L	24-hr composite	semiannually	2
Nitrobenzene	µg/L	24-hr composite	semiannually	2
Thallium	µg/L	24-hr composite	semiannually	2
Toluene	µg/L	grab	semiannually	2
Tributyltin	ng/L	24-hr composite	semiannually	2
1,1,1-Trichloroethane	µg/L	grab	semiannually	2
Acrylonitrile	µg/L	grab	semiannually	2
Aldrin	µg/L	24-hr composite	semiannually	2
Benzene	µg/L	grab	semiannually	2
Benzidine	µg/L	24-hr composite	semiannually	2
Beryllium	µg/L	24-hr composite	semiannually	2
Bis(2-chloroethyl) ether	µg/L	24-hr composite	semiannually	2
Bis(2-ethylhexyl) phthalate	µg/L	24-hr composite	semiannually	2
Carbon Tetrachloride	µg/L	grab	semiannually	2
Chlordane ¹²	µg/L	24-hr composite	semiannually	13
Chlorodibromomethane	µg/L	grab	semiannually	2
Chloroform	µg/L	grab	semiannually	2
DDT ¹²	µg/L	24-hr composite	semiannually	2
1,4-dichlorobenzene	µg/L	24-hr composite	semiannually	2
3,3'-dichlorobenzidine	µg/L	24-hr composite	semiannually	2
1,2-dichloroethane	µg/L	grab	semiannually	2
1,1-dichloroethylene	µg/L	grab	semiannually	2
Dichlorobromomethane	µg/L	grab	semiannually	2

¹³ The approved test procedures for Chlordane are methods: EPA 617, EPA 608.3 and EPA 625.1. In EPA 608.3, "In this revision to Method 608, Chlordane has been listed as the alpha- and gamma- isomers in Table 1. Reporting may be by the individual isomers, or as the sum of the concentrations of these isomers, as requested or required by a regulatory/control authority or in a permit." The 2015 Ocean Plan specifies that the sum shall be reported for compliance purposes.

Parameter	Units	Sample Type ⁶	Minimum Sampling Frequency ⁷	Required Analytical Test Method
Dichloromethane	µg/L	grab	semiannually	2
1,3-Dichloropropene	µg/L	grab	semiannually	2
Dieldrin	µg/L	24-hr composite	semiannually	2
2,4-dinitrotoluene	µg/L	24-hr composite	semiannually	2
1,2-diphenylhydrazine	µg/L	24-hr composite	semiannually	2
Halomethanes ¹²	µg/L	grab	semiannually	2
Heptachlor	µg/L	24-hr composite	semiannually	2
Heptachlor Epoxide	µg/L	24-hr composite	semiannually	2
Hexachlorobenzene	µg/L	24-hr composite	semiannually	2
Hexachlorobutadiene	µg/L	24-hr composite	semiannually	2
Hexachloroethane	µg/L	24-hr composite	semiannually	2
Isophorone	µg/L	24-hr composite	semiannually	2
N-Nitrosodimethylamine	µg/L	24-hr composite	semiannually	2
N-Nitrosodi-n-propylamine	µg/L	24-hr composite	semiannually	2
N-Nitrosodiphenylamine	µg/L	24-hr composite	semiannually	2
PAHs ¹²	µg/L	24-hr composite	semiannually	2
PCBs as Aroclors ¹²	µg/L	24-hr composite	quarterly	2
PCBs as Congeners ^{12,14,15}	µg/L	24-hr composite	quarterly	2
TCDD Equivalents ^{12,16}	µg/L	24-hr composite	quarterly	2
1,1,2,2-Tetrachloroethane	µg/L	grab	semiannually	2
Tetrachloroethylene	µg/L	grab	semiannually	2
Toxaphene	µg/L	24-hr composite	semiannually	2

¹⁴ PCBs as congeners shall be individually quantified (or quantified as mixtures of isomers of a single congener in co-elutions as appropriate) using USEPA proposed method 1668c. PCBs as congeners shall be analyzed using method EPA 1668c for three years and an alternate method may be used if none of the PCB congeners are detected for three years using method EPA 1668c.

USEPA recommends that until USEPA proposed method 1668c for PCBs is incorporated into 40 CFR § 136, Dischargers should use for discharge monitoring reports/State monitoring reports: (1) USEPA method 608 for monitoring data, reported as aroclor results, that will be used for assessing compliance with WQBELs (if applicable) and (2) USEPA proposed method 1668c for monitoring data, reported as 41 congener results, that will be used for informational purposes to help assess concentrations in the receiving water.

¹⁵ To facilitate interpretation of sediment/fish tissue data and TMDL development, PCB congeners whose analytical characteristics resemble those of PCB-18, 28, 37, 44, 49, 52, 66, 70, 74, 77, 81, 87, 99, 101, 105, 110, 114, 118, 119, 123, 126, 128, 138, 149, 151, 153, 156, 157, 158, 167, 168, 169, 170, 177, 180, 183, 187, 189, 194, 201, and 206 shall be reported as a sum and individually quantified (or quantified as mixtures of isomers of a single congener in co-elutions as appropriate).

¹⁶ USEPA Method 1613 shall be used to analyze TCDD equivalents.

Parameter	Units	Sample Type ⁶	Minimum Sampling Frequency ⁷	Required Analytical Test Method
Trichloroethylene	µg/L	grab	semiannually	2
1,1,2-Trichloroethane	µg/L	grab	semiannually	2
2,4,6-Trichlorophenol	µg/L	24-hr composite	semiannually	2
Vinyl chloride	µg/L	grab	semiannually	2
Methyl-tert-butyl-ether	µg/L	grab	semiannually	2

V. WHOLE EFFLUENT TOXICITY (WET) TESTING REQUIREMENTS

A. Chronic Toxicity Testing

1. Discharge In-stream Waste Concentration (IWC) for Chronic Toxicity

The chronic IWC is the concentration of a pollutant or the parameter toxicity in the receiving water after mixing. The chronic toxicity IWC for Discharge Point 001 is based on 1 part effluent to 108 parts seawater or 0.92 percent effluent.

2. Sample Volume and Holding Time

The total sample volume shall be determined by the specific toxicity test method used. Sufficient sample volume shall be collected to perform the required toxicity test. For the receiving water, sufficient sample volume shall also be collected during accelerated monitoring for subsequent TIE studies, if necessary, at each sampling event. All toxicity tests shall be conducted as soon as possible following sample collection. No more than 36 hours shall elapse before the conclusion of sample collection and test initiation.

3. Chronic Marine Species and Test Methods

If effluent samples are collected from outfalls discharging to receiving waters with salinity >1 ppt, the Discharger shall conduct the following chronic toxicity tests on effluent samples, at the in-stream waste concentration for the discharge, in accordance with species and test methods in Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to West Coast Marine and Estuarine Organisms (EPA/600/R-95/136, 1995). Artificial sea salts or hypersaline brine shall be used to increase sample salinity if needed. In no case shall these species be substituted with another test species unless written authorization from the Executive Officer is received.

- A static renewal toxicity test with the topsmelt, *Atherinops affinis* (Larval Survival and Growth Test Method 1006.0).
- A static non-renewal toxicity test with the red abalone, *Haliotis rufescens* (Larval Shell Development Test Method).
- A static non-renewal toxicity test with the giant kelp, *Macrocystis pyrifera* (Germination and Growth Test Method 1009.0).

4. Species Sensitivity Screening

Species sensitivity screening shall be conducted during this permit's first required sample collection. The Discharger shall collect a single effluent sample to initiate and concurrently conduct three toxicity tests using the fish, an invertebrate, and the alga species previously referenced. This sample shall also be analyzed for the parameters required on a monthly frequency for the discharge, during that given month. As allowed

under the test method for the *Atherinops affinis*, a second and third sample may be collected for use as test solution renewal water as the seven-day toxicity test progresses. If the result of all three species is "Pass" and "Percent Effect" are less than or equal to 10, at the discharge IWC, then the Discharger may continue to test with the species chosen based on the most recent, previous species sensitivity rescreening, after consultation with and written direction from the Regional Water Board staff. If only one species fails, then that species shall be used for routine monitoring during the permit cycle. Likewise, if two or more species result in "Fail", then the species that exhibits the highest "Percent Effect" at the discharge IWC during the suite of species sensitivity screening shall be used for routine monitoring during the permit cycle, until such time as a rescreening is required.

Species sensitivity rescreening is required every 24 months if there has been discharge during dry weather conditions. If the discharge is intermittent and occurs only during wet weather, rescreening is not required. If rescreening is necessary, the Discharger shall rescreen with the marine vertebrate species, a marine invertebrate species, and the alga species previously referenced, and continue to monitor with the most sensitive species. If the first suite of rescreening tests demonstrates that the same species is the most sensitive then the rescreening does not need to include more than one suite of tests. If a different species is the most sensitive or if there is ambiguity, then the Discharger may proceed with suites of screening tests for a minimum of three, but not to exceed five suites.

During the calendar month, toxicity tests used to determine the most sensitive test species shall be reported as effluent compliance monitoring results for the chronic toxicity MDEL.

5. Quality Assurance and Additional Requirements

Quality assurance measures, instructions, and other recommendations and requirements are found in the test methods manual previously referenced. Additional requirements are specified below.

- a. The discharge is subject to determination of "Pass" or "Fail" from a chronic toxicity test using the Test of Significant Toxicity statistical t-test approach described in the National Pollutant Discharge Elimination System Test of Significant Toxicity Implementation Document (EPA 833-R-10-003, 2010), Appendix A, Figure A-1, and Table A-1, and Appendix B, Table B-1. The null hypothesis (H_0) for the TST statistical approach is: Mean discharge IWC response $\leq 0.75 \times$ Mean control response. A test result that rejects this null hypothesis is reported as "Pass." A test result that does not reject this null hypothesis is reported as "Fail." The relative "Percent Effect" at the discharge IWC is defined and reported for each toxicity test as: $((\text{Mean control response} - \text{Mean discharge IWC response}) \div \text{Mean control response}) \times 100$. This is a t-test (formally Student's t-Test), a statistical analysis comparing two sets of replicate observations – in the case of WET, only two test concentrations (i.e. a control and IWC). The purpose of this statistical test is to determine if the means of the two sets of observations are different (i.e. if the IWC or receiving water concentration differs from the control (the test result is "Pass" or "Fail")). The Welch's t-test employed by the TST statistical approach is an adaptation of Student's t-test and is used with two samples having unequal variances.
- b. If the effluent toxicity test does not meet all test acceptability criteria (TAC) specified in the referenced test method Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to West Coast Marine and Estuarine

Organisms (EPA/600/R-95/136, 1995) (see Table E-6, below), then the Discharger must re-sample and re-test within 14 days.

Table E-6. USEPA Test Methods and Test Acceptability Criteria

Species & USEPA Test Method Number	Test Acceptability Criteria (TAC)
Topsmelt, <i>Atherinops affinis</i> , Larval Survival and Growth Test Method 1006.01. (Table 3 of Test Method)	80% or greater survival in controls; average dry weight per surviving organism in control chambers equals or exceeds 0.85 mg. LC50 with copper must be $\leq 205 \mu\text{g/L}$, <25% MSD for survival and <50% MSD for growth. If the test starts with 9 day old larvae, the mean weight per larva must exceed 0.85 milligrams in the reference and brine controls; the mean weight of preserved larvae must exceed 0.72 milligrams. (required)
Purple Sea Urchin, <i>Strongylocentrotus purpuratus</i> , and the Sand Dollar, <i>Dendraster excentricus</i> , Fertilization Test Method 1008.0 (Table 7 of Test Method)	70% or greater egg fertilization in controls, must achieve a MSD of <25%, and appropriate sperm counts. (required)
Red Abalone, <i>Haliotis rufescens</i> , Larval Shell Development Test Method (Table 3 of Test Method)	80% or greater normal shell development in the controls; must have statistical significant effect at $56 \mu\text{g/L}$ zinc and achieve a MSD of <20%. (required)
Giant Kelp, <i>Macrocystis pyrifera</i> , Germination and Growth Test Method 1009.0 (Table 3 of Test Method)	70% or greater germination in controls, $\geq 10 \mu\text{m}$ germ-tube length in controls, No Observed Effect Concentration (NOEC) must be below $35 \mu\text{g/L}$ in the reference toxicant test, and must achieve a MSD of <20% for both germination and germ-tube length in the reference toxicant. (required)

- c. Dilution water and control water, including brine controls, shall be 1- μm -filtered uncontaminated natural seawater, hypersaline brine prepared using uncontaminated natural seawater, or laboratory water prepared and used as specified in the test methods manual. If dilution water and control water is different from test organism culture water, then a second control using culture water shall also be used.
- d. Monthly reference toxicant testing is sufficient. All reference toxicant test results should be reviewed and reported using the EC25¹⁷.

¹⁷ EC25 is a point estimate of the toxicant concentration that would cause an observable adverse effect (e.g. death, immobilization, or serious incapacitation) in 25 percent of the test organisms.

- e. The Discharger shall perform toxicity tests on final effluent samples. Chlorine and ammonia shall not be removed from the effluent sample prior to toxicity testing, unless explicitly authorized under this section of the MRP and the rationale is explained in the Fact Sheet (Attachment F).

6. Preparation of an Initial Investigation Toxicity Reduction Evaluation (TRE) Work Plan

The Discharger shall prepare and submit a copy of the Discharger's initial investigation TRE work plan to the Executive Officer of the Regional Water Board for approval within 90 days of the effective date of this permit. If the Executive Officer does not disapprove the work plan within 60 days, the work plan shall become effective. The Discharger shall use USEPA manual EPA/833B-99/002 (municipal) as guidance, or the most current version. This work plan shall describe the steps that the Discharger intends to follow if toxicity is detected. At a minimum, the TRE Work Plan must contain the provisions in Attachment G. This work plan shall describe the steps that the Discharger intends to follow if toxicity is detected. At a minimum the work plan shall include:

- a. A description of the investigation and evaluation techniques that will be used to identify potential causes and sources of toxicity, effluent variability, and treatment system efficiency.
- b. A description of the Facility's methods of maximizing in-house treatment efficiency and good housekeeping practices, and a list of all chemicals used in the operation of the Facility; and,
- c. If a TIE is necessary, an indication of the person who would conduct the TIEs (i.e., an in-house expert or an outside contractor).

7. Accelerated Monitoring Schedule for Maximum Daily Single Result: "Fail."

The Maximum Daily single result shall be used to determine if accelerated testing needs to be conducted.

Once the Discharger becomes aware of this result, the Discharger shall implement an accelerated monitoring schedule within 5 calendar days of the receipt of the result. However, if the sample is contracted out to a commercial laboratory, the Discharger shall ensure that the first of four accelerated monitoring tests is initiated within seven calendar days of the Discharger becoming aware of the result. The accelerated monitoring schedule shall consist of four toxicity tests (including the discharge IWC), conducted at approximately two-week intervals, over an eight-week period; in preparation for the TRE process and associated reporting, these results shall also be reported using the EC25. If each of the accelerated toxicity tests results in "Pass," the Discharger shall return to routine monitoring for the next monitoring period. If one of the accelerated toxicity tests results in "Fail," the Discharger shall immediately implement the TRE Process conditions set forth below. During accelerated monitoring schedules, only TST results ("Pass" or "Fail") for chronic toxicity tests shall be reported as effluent compliance monitoring results for the chronic toxicity MDEL.

8. Toxicity Reduction Evaluation (TRE) Process

During the TRE Process, monthly effluent monitoring shall resume and TST results ("Pass" or "Fail") for chronic toxicity tests shall be reported as effluent compliance monitoring results for the chronic toxicity MDEL.

- a. Preparation and Implementation of Detailed TRE Work Plan. The Discharger shall immediately initiate a TRE using, according to the type of treatment facility, USEPA manual Toxicity Reduction Evaluation Guidance for Municipal Wastewater Treatment Plants (EPA/833/B-99/002, 1999) and, within 15 days, submit to the

Executive Officer a Detailed TRE Work Plan, which shall follow the generic Initial Investigation TRE Work Plan revised as appropriate for this toxicity event. It shall include the following information, and comply with additional conditions set by the Executive Officer:

- i. Further actions by the Discharger to investigate, identify, and correct the causes of toxicity.
 - ii. Actions the Discharger will take to mitigate the effects of the discharge and prevent the recurrence of toxicity.
 - iii. A schedule for these actions, progress reports, and the final report.
- b. TIE Implementation. The Discharger may initiate a TIE as part of a TRE to identify the causes of toxicity using the same species and test method and, as guidance, USEPA manuals: Methods for Aquatic Toxicity Identification Evaluations: Phase I Toxicity Characterization Procedures (EPA/600/6-91/003, 1991); Methods for Aquatic Toxicity Identification Evaluations, Phase II Toxicity Identification Procedures for Samples Exhibiting Acute and Chronic Toxicity (EPA/600/R-92/080, 1993); Methods for Aquatic Toxicity Identification Evaluations, Phase III Toxicity Confirmation Procedures for Samples Exhibiting Acute and Chronic Toxicity (EPA/600/R-92/081, 1993); and Marine Toxicity Identification Evaluation (TIE): Phase I Guidance Document (EPA/600/R-96-054, 1996). The TIE should be conducted on the species demonstrating the most sensitive toxicity response.
- c. Many recommended TRE elements parallel required or recommended efforts for source control, pollution prevention, and storm water control programs. TRE efforts should be coordinated with such efforts. As toxic substances are identified or characterized, the Discharger shall continue the TRE by determining the sources and evaluating alternative strategies for reducing or eliminating the substances from the discharge. All reasonable steps shall be taken to reduce toxicity to levels consistent with toxicity evaluation parameters.
- d. The Discharger shall continue to conduct routine effluent monitoring for compliance determination purposes while the TIE and/or TRE is taking place. Additional accelerated monitoring and TRE work plans are not required once a TRE has begun.
- e. The Regional Water Board recognizes that toxicity may be episodic and identification of causes and reduction of sources of toxicity may not be successful in all cases. The TRE may be ended at any stage if monitoring finds there is no longer toxicity.

9. Reporting

The Self-Monitoring Report (SMR) shall include a full laboratory report for each toxicity test. This report shall be prepared using the format and content of the test methods manual chapter called Report Preparation, and shall include:

- a. The valid toxicity test results for the TST statistical approach, reported as "Pass" or "Fail" and "Percent Effect" at the chronic toxicity IWC for the discharge. All toxicity test results (whether identified as valid or otherwise) conducted during the calendar month shall be reported on the SMR due date specified in Table E-16.
- b. Summary water quality measurements for each toxicity test (e.g. pH, dissolved oxygen, temperature, conductivity, hardness, salinity, chlorine, ammonia).

- c. The statistical analysis used in National Pollutant Discharge Elimination System Test of Significant Toxicity Implementation Document (EPA 833-R-10-003, 2010) Appendix A, Figure A-1 and Table A-1, and Appendix B, Table B-1.
- d. TRE/TIE results. The Executive Officer shall be notified no later than 30 days from completion of each aspect of TRE/TIE analyses. Prior to completion of the final TIE/TRE report, the Discharger shall provide status updates in the monthly monitoring reports, indicating which TIE/TRE steps are underway and which steps have been completed.
- e. Statistical program (e.g., TST calculator, Comprehensive Environmental Toxicity Information System, etc.) output results, including graphical plots, for each toxicity test.
- f. Graphical plots and tabular data clearly showing the laboratory's performance of the reference toxicant, for each solution, for the previous 20 tests and the laboratory's performance of the control mean, control standard deviation, and control coefficient of variation, for each solution, for the previous 12-month period.
- g. Any additional QA/QC documentation or any additional chronic toxicity-related information, upon written request of the Regional Water Board Chief Deputy Executive Officer or Executive Officer.

B. Ammonia Removal

1. Except with prior approval from the Executive Officer of the Regional Water Board, ammonia shall not be removed from bioassay samples. The Discharger must demonstrate the effluent toxicity is caused by ammonia because of increasing test pH when conducting the toxicity test. It is important to distinguish the potential toxic effects of ammonia from other pH sensitive chemicals, such as certain heavy metals, sulfide, and cyanide.
 - a. The following may be steps to demonstrate that the toxicity is caused by ammonia and no other toxicants before the Executive Officer would allow for control of pH in the test. There is consistent toxicity in the effluent and the maximum pH in the toxicity test is in the range to cause toxicity due to increased pH.
 - b. Chronic ammonia concentrations in the effluent are greater than 4 mg/L total ammonia.
 - c. Conduct graduated pH tests as specified in the toxicity identification evaluation methods. For example, mortality should be higher at pH 8 and lower at pH 6.
 - d. Treat the effluent with a zeolite column to remove ammonia. Mortality in the zeolite treated effluent should be lower than the non-zeolite treated effluent. Then add ammonia back to the zeolite-treated samples to confirm toxicity due to ammonia.
2. When it has been demonstrated that toxicity is due to ammonia because of increasing test pH, pH may be controlled using appropriate procedures which do not significantly alter the nature of the effluent, after submitting a written request to the Regional Water Board, and receiving written permission expressing approval from the Executive Officer of the Regional Water Board.

C. Chlorine Removal

Chlorine may be removed from the OWTP effluent bioassay sampled from EFF-001B because there are no appropriate sampling locations that reflect dechlorinated conditions at the outfall.

VI. LAND DISCHARGE MONITORING REQUIREMENTS – NOT APPLICABLE

VII. RECYCLING REQUIREMENTS

The reuse of the reclaimed water is regulated under a separate WDRs and Water Recycling Requirements (WRRs) for City of Oxnard Groundwater Recovery, Enhancement, and Treatment Program – Non-Potable Reuse Phase 1 Project (GREAT Program – Phase 1 Project), Order No. R4-2008-0083 as amended by Order No. R4-2011-0079, Order No. R4-2011-0079-A01, and Order No. R4-2011-0079-A02, File No. 64-104 and File No. 08-070, CI-9456.

VIII. RECEIVING WATER MONITORING REQUIREMENTS

The City of Oxnard has been monitoring receiving water at Discharge Point 001 since 1999. Local receiving water trends over the 10-kilometer sample grid have generally remained steady and the 20-year record of data shows the Oxnard discharge plume is not sufficiently large to impact regional trends. Based on the most recent information from the federally-funded Southern California Coastal Ocean Observing System (SCCOOS), the Central Bight Cooperative Water Quality Survey is complete and no additional water quality monitoring is required at this time. Due to this reduction in regional water quality monitoring, this Order reduces the area sampled to improve information on the impact of the outfall and more accurately characterize the dilution which takes place at the outfall. However, the City of Oxnard shall continue to participate on the Central Bight Cooperative Water Quality Survey steering committee and provide assistance should this cooperative program undertake analysis of the collected data to contribute to a regional understanding of seasonal patterns in water column structure.

Receiving water shall continue to be monitored for local offshore water quality, shoreline bacteria, infaunal benthic community, sediment chemistry and toxicity, regional benthic health, local fish and macroinvertebrate population and tissue, regional fish and macroinvertebrate health, local seafood safety, regional seafood safety, and kelp bed health. If annual reporting and comparative studies identify unanticipated water quality trends at the smaller one-kilometer grid size, the wider sample grid may be reinstated. All receiving water stations shall be located by state-of-the-art navigational methods (e.g. Differential Global Positioning System or DGPS); other means (e.g. visual triangulation, fathometer readings) may be used to improve the accuracy of locating stations. Water quality measurements are made with a Conductivity, Temperature and Depth Instrument (CTD), which also measures other parameters such as pH and light transmissivity.

Table E-7. Schedule for Receiving Water Monitoring Requirements

Monitoring	Number	Type	R4-2013-0094 Frequency	R4-2018-0140 Frequency	Reporting
Local offshore water quality	25	CTD, ammonia, bacteria	--	Quarterly	Annually
Regional offshore water quality	48	CTD and SCCWRP support	Quarterly	---	--
Bacteria/ammonia offshore water quality	18/ three depths	Bacteria, ammonia	Quarterly	--	--
Shoreline	9	Bacteria	Quarterly	Quarterly	Annually

Monitoring	Number	Type	R4-2013-0094 Frequency	R4-2018-0140 Frequency	Reporting
Local Benthic Infaunal community	7	Community and population	Annually	Year two	Year three
Local Sediment chemistry	7	Chemistry	Annually	Year two	Year two
Sediment toxicity	2	Amphipod	Annually	Year two	Year two
Regional Benthic	SCCWRP support only		2013	2018, 2023	SCCWRP
Local fish macroinvertebrate	3	Population	Annually	Year two	Year two
Local fish macroinvertebrate	3	Tissue	Annually	Year two	Year two
Local bagged mussels	3	Tissue	Special Study	Year four	Year four
Regional fish and macroinvertebrate	SCCWRP support only		2013	2018, 2023	SCCWRP
Local seafood safety	5 species in 3 zones	Test against consumption advisories	Year one, three and five	Year four	Year four
Regional seafood safety	SCCWRP support only		2013	2018, 2023	SCCWRP
Kelp bed monitoring	SCCWRP support only		2013	2018, 2023	SCCWRP

A. Water Quality Monitoring

Water quality monitoring is designed to determine if Ocean Plan and Basin Plan objectives for physical and chemical parameters and bacteria are being met. Water quality data will be collected to provide the information necessary to demonstrate compliance with the water quality standards and to contribute to the Bight'18 and Bight'23 regional monitoring programs, led by SCCWRP. Sample collection for water quality monitoring shall follow protocols described in the most current edition of the *Field Operations Manual for Marine Water-Column, Benthic, and Trawl Monitoring in Southern California, SCCWRP*. Water quality data shall be analyzed to estimate spatial extent and movement of the wastewater plume over time. Changes in monitoring measurements shall be quantified each year, contrasted with the previous five years, and summarized annually.

1. Inshore/Offshore Monitoring - The Discharger shall monitor receiving water quality quarterly at 25 new core Receiving Water Column Monitoring Stations from A001 to E005 (Table E-2 and Attachment B-4) as follows.

Table E-8. Offshore Receiving Water Monitoring Requirements

Parameter	Units	Sample Type	Minimum Sampling Frequency	Required Analytical Test Method
Dissolved oxygen	mg/L	continuous profile	quarterly	2
Water temperature	°C	continuous profile	quarterly	2
Light transmittance	% transmittance	continuous profile	quarterly	18
Salinity	ppt	continuous profile	quarterly	2

¹⁸ Light transmittance (transmissivity) shall be measured with a transmissometer, using equipment and procedure similar to that described by L.V. Whitney [*Transmission of Solar Energy and the Scattering Produced by Suspensoids in Lake Waters*, Transactions of the Wisconsin Academy of Sciences, Arts, and Letters, Vol. 31 (1938)]. Results shall be expressed as the percent of light transmittance. Path length of transmissometer should be noted.

Parameter	Units	Sample Type	Minimum Sampling Frequency	Required Analytical Test Method
pH	pH units	continuous profile	quarterly	2
Chlorophyll a	µg/L	continuous profile	quarterly	2
Visual observations	---	---	quarterly	19
Total coliform	MPN or CFU/100 mL	grab, surface and mid-depth and near bottom ²⁰	quarterly	2
Fecal coliform	MPN or CFU/100 mL	grab, surface and mid-depth and near bottom ²⁰	quarterly	2
<i>Enterococcus</i>	MPN or CFU/100 mL	grab, surface and mid-depth and near bottom ²⁰	quarterly	2
Ammonia nitrogen	mg/L	grab, surface and mid-depth and near bottom ²⁰	quarterly	2

Concurrent with the CTD profiling survey, discrete samples shall be collected quarterly for ammonia and fecal coliform, total coliform and *Enterococcus* at the surface, mid depth and near the bottom (or as deep as practicable for those stations located in depths less than 45 m). Sampling for bacteria is necessary to ensure the bacteria discharged at the outfall do not survive marine conditions and do not contribute to the bacteria identified by shoreline monitoring.

2. Shoreline Monitoring - The Discharger shall monitor receiving water quality at nine Ventura County Shoreline Bacteriological Monitoring Stations listed in Attachment B-8 for the constituents that follow:

¹⁹ Observations of wind (direction and speed), weather (e.g., cloudy, sunny, or rainy), current (e.g., direction), and tidal conditions (e.g., high or low tide) shall be made and recorded (every four hours during offshore sampling) at the time samples of the waters of the Pacific Ocean (shore, nearshore, and all offshore stations) are collected.

Observations of water color, discoloration, oil and grease, turbidity, odor, materials of sewage origin in the water or on the beach, and unusual or abnormal amounts of floating or suspended matter in the water or on the beach, rocks and jetties, or beach structures shall also be made and recorded at stations or while in transit. The character and extent of such matter shall be described. The dates, times and depths of sampling and these observations shall also be reported.

²⁰ Bottom sampling shall be done 2.0 m (6.6 ft) above the seabed.

Table E-9. Shoreline Receiving Water Monitoring Requirements for Bacteria

Parameter	Units	Sample Type	Minimum Sampling Frequency	Required Analytical Test Method
Total coliform	MPN or CFU/100 mL	grab	quarterly	2
Fecal coliform	MPN or CFU/100 mL	grab	quarterly	2
<i>Enterococcus</i>	MPN or CFU/100 mL	grab	quarterly	2

Visual observations shall be recorded when bacteriological samples are collected. Monitoring at these nine stations is conducted for the purposes of public health assessment and to ensure protection for public recreational use of coastal ocean waters, and the results are not intended for use as compliance sites for OWTP, unless offshore bacteria monitoring demonstrates the outfall bacteria persist to the shoreline. In the event of stormy weather that makes sampling hazardous or impractical, these samples can be rescheduled.

B. Benthic Monitoring

Benthic monitoring includes infauna and sediment sampling. The data collected are used for the regular assessment of trends in sediment contamination and biological response along a fixed grid of sites within the influence of the discharge.

1. Local Benthic Monitoring - This survey is designed to determine if benthic conditions under the influence of the discharge are changing over time.

The Discharger shall monitor infauna, sediment chemistry and sediment toxicity at 7 receiving water benthic monitoring stations of RWS-001 to RWS-007 in year two of the permit and report changes in the conditions as compared to the historic and the most recent receiving water reports (See Table E-3 and Attachment B-5) in the year three annual report.

- a. Infaunal Community Survey - These stations shall be sampled for benthic infauna²¹ during late summer (August/ September). Bottom samples for benthic infaunal analyses shall be taken at each benthic station prior to trawl sampling. The following determinations shall be made at each station, where appropriate;
 - i. Identification of all organisms to lowest possible taxon (usually species); and,
 - ii. Total biomass of:
 - (a) Mollusks;
 - (b) Echinoderms;
 - (c) Annelids/polychaetes;
 - (d) Crustaceans; and,
 - (e) All other macroinvertebrates.

²¹ These bottom samples shall be taken by means of a 0.1 m² (1.1 ft²) modified Van Veen sediment grab sampler. The entire contents of each sample shall be passed through a 1.0 mm (0.039 in.) mesh screen to retrieve the benthic organisms. These organisms shall be fixed in 10% buffered formalin and transferred to 70% ethanol within two to seven days for storage. All specimens retrieved shall be archived.

- iii. Community structure²² analysis for benthic infauna for each station and each replicate. Mean, median, range, standard deviation, and 95% confidence limits, if appropriate, for values determined above. The Discharger may use other statistical tools sufficient to determine temporal and spatial trends in the marine environment.
- b. Sediment Chemistry Survey - Testing shall be required for these parameters during late summer (August/ September). Bottom samples for sediment chemistry analyses shall be taken at each benthic station prior to trawl sampling. All benthic sediment samples shall be taken at each station by means of a 0.1 m² (1.1 ft²) modified Van Veen sediment grab sampler. Sub-samples (upper two centimeters) of sediment from each sample shall be collected and analyzed separately for the following parameters at each station:
 - i. Total organic carbon (TOC) (mg/kg dry wt);
 - ii. Dissolved sulfides (water soluble) (mg/kg dry wt);
 - iii. Total Kjeldahl nitrogen (mg/kg dry wt);
 - iv. Grain size (sufficiently detailed to calculate percent weight in relation to phi size); and,
 - v. Arsenic; Cadmium; Chromium (total); Copper; Lead; Mercury; Nickel; Silver; Zinc; Cyanide; Phenolic compounds (non-chlorinated); Phenolic compounds (chlorinated); Total halogenated organic compounds; Aldrin and Dieldrin; Endrin; HCH; Chlordane and related compounds; Total DDT; DDT derivatives; Total PCB; PCB derivatives; Toxaphene; Total PAH; PAH derivatives. The data for these parameters shall be expressed in µg/kg dry weight. Additional testing shall be conducted such that a full priority pollutant scans is performed on the sediment samples.
- c. Chronic Sediment Toxicity Survey - Sediment toxicity testing shall be conducted in August/September at the two receiving water sediment monitoring stations RWS-003 and RWS-007. Three replicate samples shall be collected for testing at each station. Sub-samples (upper two centimeters) shall be taken from each sediment sample and tested for chronic toxicity. Testing shall be conducted using one of the three amphipod species *Eohaustorius estuarius*, *Leptocheirus plumulosus*, and *Rhepoxynius abronius* in accordance with EPA 600/R-94/0925 (USEPA, 1994), *Methods for Assessing the Toxicity of Sediment-associated Contaminants with Estuarine and Marine Amphipods*, and the *Southern California Bight Project sediment toxicity testing guidelines* (Bight '18 Toxicology Committee, 2018). Test results shall be reported in percent survival, assessed for the presence of persistent toxicity, and included in the annual monitoring report. If persistent toxicity is observed at a sediment sampling location, a Phase I Toxicity Identification Evaluation (TIE) shall be conducted as defined in the *Sediment Toxicity Identification (TIE) Phase I, II, and III Guidance Document* (EPA/R-07/080). The Discharger shall submit a Sediment Toxicity TIE Work Plan within 90 days of the effective date of this

²² Community structure analysis of benthic infauna shall include number of species, number of individuals per species, total numerical abundance, species abundance per grab, species richness, species diversity (e.g., Shannon-Wiener), species evenness and dominance per station and replicate, similarity analyses (e.g., Bray-Curtis, Jaccard or Sorensen), cluster analyses (using unweighted pair-group method) or other appropriate multivariate statistical techniques approved by the Executive Officer of this Regional Water Board and the Infaunal Index.

Order. The work plan shall define persistent toxicity and outline the procedures that will take place if persistent toxicity is observed.

2. Regional Benthic Monitoring - This regional survey is designed to determine the extent, distribution, magnitude and trend of ecological change in soft-bottom benthic habitats within the Southern California Bight and the relationship between biological response and contaminant exposure. The data collected will be used to assess the condition of the sea-floor environment and the health of the biological resources within the Bight.

A regional survey of sediment chemistry and benthic infauna within the Southern California Bight took place in 2013 (Bight'13). The final survey design was determined cooperatively by the participants represented on the Regional Steering Committee. The Discharger provided support to the Bight'13 benthic surveys in the following ways:

- a. Participation on the Steering Committee
- b. Participation on the relevant Technical Committees (e.g., Information Management, Field Methods and Logistics, Benthos and Chemistry)
- c. Field sampling at sea
- d. Infaunal sample analysis
- e. Sediment chemistry analysis
- f. Data management

The Discharger's level of participation in previous Bight surveys has been consistent. The same level of participation is expected in Bight'18 and Bight'23 surveys.

C. Fish and Invertebrate Monitoring

1. Local Fish and Macroinvertebrate Survey – This survey is designed to determine how the health of demersal fish and epibenthic invertebrate communities near the discharge point is changing over time. The data collected will be used for regular assessment of temporal trends in community structure and bioaccumulation along an array of sites within the influence of the discharge. Trash and debris data will also be collected to contribute to the Santa Monica Bay Restoration Project's (SMBRP's) Sources and Loadings Program. The Discharger shall monitor fish and macroinvertebrate at three receiving water trawling stations RWT-001, RWT-002 and RWT-003 in year two (See Table E-3). Mussels will be deployed for three months (September to December) at three fixed locations for bioaccumulation (see Attachment B-7) in year four. The monitoring requirements are as follows:

- a. Local Fish and Macroinvertebrate Population Survey
 - i. Offshore trawling will occur in year two during August/September for demersal fish and epibenthic macroinvertebrates with trawl locations along sample lines A001 to A005 through E001 to E005 (see Table E-2 and Attachment B-7).
 - ii. Trawling methods shall follow the protocols described in the most current edition of the *Field Operations Manual for Marine Water-Column, Benthic, and Trawl Monitoring in Southern California, SCCWRP*.